



Preservation of human skin integrity and metabolism during 10 days in culture with NativeSkin[®] system

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Human epidermis is not vascularized and oxygen only reaches epidermal cells by diffusing through the stratum corneum or from the dermal papillae which contains a dense capillary network. It is known that the main bioenergetic pathway of human epidermis is anaerobic glycolysis (Ronquist et al., 2003). Therefore, glucose consumption and a high production of lactate indicate an active metabolism in skin. This pathway has not been well investigated in human skin biopsies cultured *ex vivo*.

Genoskin has developed NativeSkin[®], a standardized *ex vivo* human skin model containing both epidermis and dermis, produced from surgical skin samples. The skin model is embedded in a proprietary nourishing gel-like matrix with epidermal surface left in direct contact with the air. NativeSkin[®] represents a highly predictive and cost-effective last-line screening tool in laboratory conditions prior to *in vivo* clinical evaluations. We study here the integrity, viability, and metabolism of NativeSkin® models after 10 days of ex vivo culture.

Experimental Workflow



Donor 3

NATIVESKIN[®] MODEL CULTURE DURING 10 DAYS AND SAMPLING.

Donor 2

The experiment was performed on 7 donors. NativeSkin[®] M models (15 mm of diameter) were cultured for 10 days. Culture media was changed dail At Day 0 and Day 10, skin models were sampled as described. Culture media was frozen at -80°C.

	Donor #	Gender	Age	Anatomical site	Number of replicates
lv –	1		42		n=2
' y .	2		28		n=2
	3		58		n=4
	4	Female	37	Abdomen	n=4
	5		37		n=4
	6		38		n=2
	7		52		n=2

NativeSkin[®] shows cell proliferation and no apoptosis at day 10



ANTI-KI67 AND ANTI-ACTIVE CASPASE-3 IMMUNOSTAININGS AND QUANTIFICATION.

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Immunostainings were performed on 5µm thick skin slices. Representative images at Day 0 and Day 10 are presented. Quantifications were performed on 6 representative pictures of the epidermis.

NativeSkin[®] maintains normal skin integrity and a significant cell viability at day 10

Donor 5

Donor 6





Donor 4

NORMAL SKIN INTEGRITY AFTER 10 DAYS OF EX VIVO CULTURE.

Hematoxylin and Eosin staining was performed on 5µm thick skin slices. A representative image of NativeSkin[®] model at Day 0 and Day 10 is presented for each donor.



CELL VIABILITY AFTER 10 DAYS EX VIVO CULTURE

WST-8 assay was performed on small punch biopsies (2 in the center and 4 in the edge), in order to assess cell viability in different area of the model.

NativeSkin[®] consumes glucose and produces lactate at day 10



LACTATE PRODUCTION AND GLUCOSE **CONSUMPTION IN CULTURE MEDIA.** Glucose consumption Concentrations of lactate produced and glucose consumed in culture media between Day 9 and Day 10 were dosed. The bars represent the mean ± SD of replicates.

LACTATE PRODUCTION AND GLUCOSE

Donor 7

NativeSkin[®] maintains normal ROS concentration at day 10

Center of the biopsy





ROS CONCENTRATION IN NATIVESKIN® MODELS H₂O₂ concentration in NativeSkin[®] cultured 10 days was measured on 1 frozen biopsy for each sample using ROS-Glo[™] H₂O₂ Assay (Promega). The bars represent the mean +/- SD of replicates.

NativeSkin[®] is an ex vivo human skin model that can normally be maintained in standard culture conditions up to 7 days when shipped to customers. Here, we demonstrate for the first time maintaining the skin model 10 days in ex vivo culture while preserving normal skin integrity, significant cell viability, proliferative cells and low levels of apoptosis. We demonstrate that the skin metabolism of NativeSkin[®] at 10 days is comparable to fresh skin by measuring glucose consumption and lactate production using Promega's Glucose-Glo[™] and Lactate-Glo[™] Assays. We also demonstrate that NativeSkin[®] maintains normal ROS concentration after 10 days of *ex vivo* culture, using Promega's ROS-Glo[™] H₂O₂ Assay. Our next step will be to evaluate the activity of specific human skin cytochromes involved in metabolization of drugs topically applied on NativeSkin[®].