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FOR IN VITRO TESTING

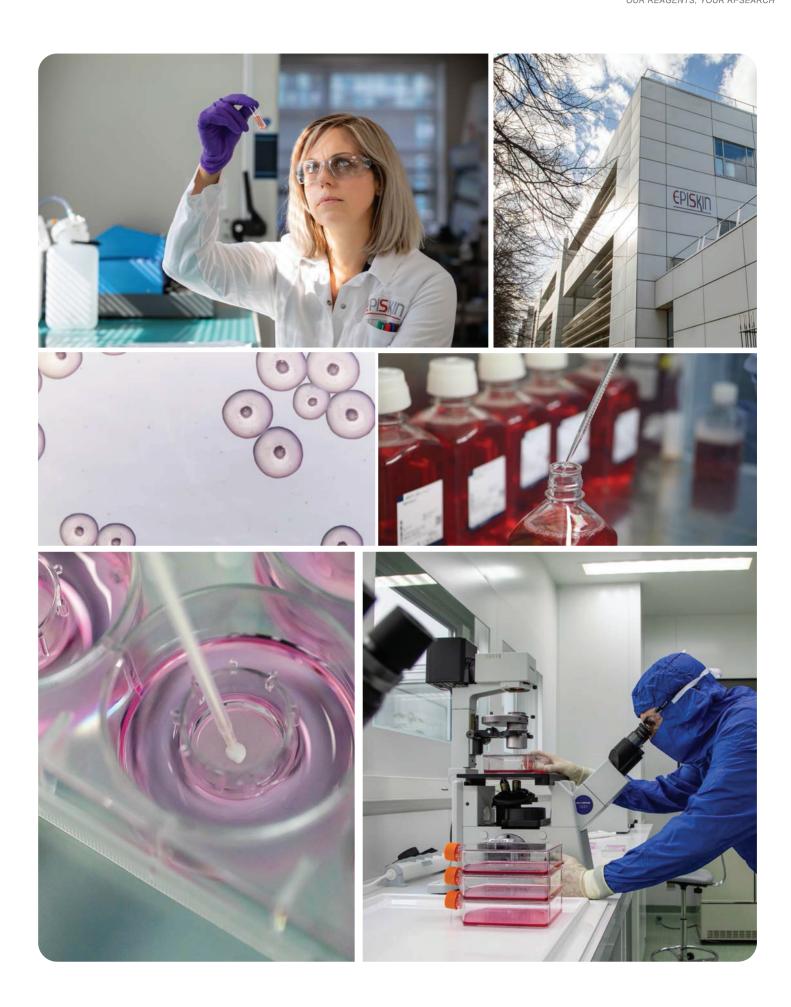


APPLICATIONS

Skin irritation					
Skin corrosion					
UV exposure					
Bacterial adhesion					
DNA Damage					
Omics					
Permeability					
Eye irritation					
Medical Devices					
Microbiome					
Skin immune response					
Pigmentation/Depigmentation					
Oral & Gingival care					
Oesophageal Irritation					
Bladder irritation					
Vaginal irritation					

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EPISKIN: your reliable and trustworthy partner for your benefits

We innovate

since 1992 for a better science in producing advanced, relevant and robust in vitro 3D models allowing our users to predict human response. Very strict and unique quality controls help us to supply unmatched quality models. Our innovation, your science.

We dedicate

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. Worldwide

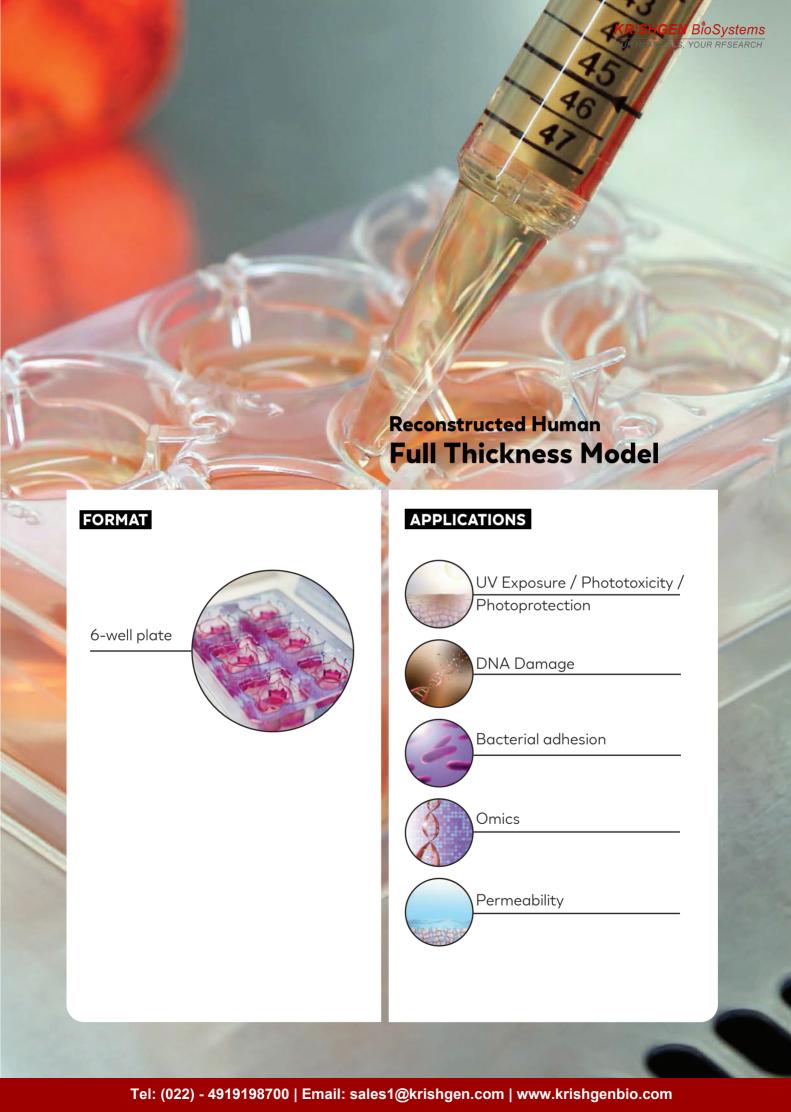
We deliver

Innovation

Expertise

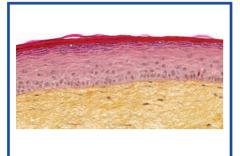
worldwide, serving all customers with our 3 partners in India, Korea and Japan and with our 2 subsidiaries in Brazil and China and our headquarters in France. You order, we deliver.

ADVANCED 3D MODELS FOR A BETTER,
MORE PREDICTIVE AND MORE ETHICAL SCIENCE





Reconstructed Human Full Thickness Model



NAME

T-Skin™ / Reconstructed Human Full Thickness Skin Model

DESCRIPTION

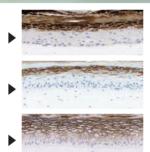
T-Skin[™] is an in vitro reconstructed skin which consists of a dermal equivalent with human fibroblasts overlaid by a stratified, well differentiated epidermis derived from normal human keratinocytes cultured on an inert polycarbonate filter.

This model exists at different stages of maturity.

SPECIFIC MARKERS

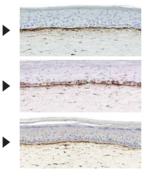
Differentiation markers:

- Filaggrin Involucrin Loricrin
- Cvtokeratin 10
- Transglutaminase-1



Dermal-Epidermal junction markers

- Collagen IV, VII, XII
- Laminin V
- Perlecan BP antigen



Proliferation markers

Ki67



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Reconstructed skin to create in vitro flexible models of skin aging: new results and prospects.

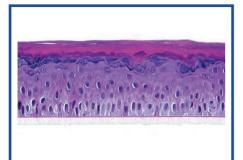
Asselineau D, Ricois S, Pageon H, Zucchi H, Girardeau-Hubert S, Deneuville C, Haydont V, Neiveyans V, Lorthois I, In Farage MA, Miller W, Maibach H, editors. Springer Berlin Heidelberg Textbook of Áging Skin p 1203-1228.

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Reconstructed Human **Epidermis**



NAME

SkinEthic[™] RHE / Reconstructed Human Epidermis

DESCRIPTION

SkinEthicTM RHE is an *in vitro* reconstructed human epidermis from normal human keratinocytes cultured on an inert polycarbonate filter at the air-liquid interface. It is histologically similar to the *in vivo* human epidermis.

Our strong believe in Science and our continuous improvement with ISO 9001 certification push us to keep improving the production process of our model: From cell extraction to reconstruction with chemically defined biocomponents and **medium.**

Every single biocomponent of each step of our production is clearly defined and their traceability is guaranteed. The process is then more secured, allowing to deliver a SkinEthicTM RHE model more reproducible, robust and reliable than ever.

Different maturities and surfaces are available.

SPECIFIC MARKERS

Differentiation markers:

- Filaggrin
- Involucrin Loricrin
- Transglutaminase-1
 Keratin 10
 Keratin 5
 CD44







Presence of different epidermal classes of lipids comprising ceramides

Dermal-Epidermal junction markers

Type IV collagen

► Laminin V Alpha6Beta4-integrin BP antigen



Proliferation markers

Ki67



are epidermal models on an innovative insert. Epidermal or mucosal reconstructed on this insert are histologically similar to *in vivo* human epidermis and mucosa.

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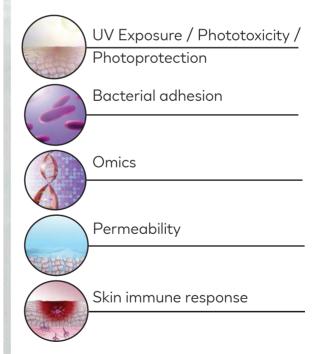


Reconstructed Human Epidermis with Langerhans Cells

FORMAT

0.5 cm²

APPLICATIONS





Reconstructed Human Epidermis with Langerhans Cells



NAME

SkinEthic™ RHE-LC / Human Epidermal Model Langerhans

DESCRIPTION

The SkinEthic[™] RHE-LC model is a standard epidermal model in which Langerhans cells progenitors have been integrated. During the tissue reconstruction, these immature cells have differentiated into antigen-presenting Langerhans cells expressing the specific marker CD207 (langerin). They are mostly located and evenly spread within the supra-basal epidermal layer. This model is therefore expected to be a useful tool

for skin immune response studies.

SPECIFIC MARKERS

Langerhans cells markers:

- CD207 Langerin
- Birbeck granules

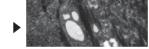
CD1

CD80

CD86

CCR7





Differentiation markers:

Filaggrin

Involucrin

Loricrin

Keratin 10

Keratin 5

Presence of different epidermal classes of lipids comprising ceramides

Dermal-Epidermal junction markers

Type IV collagen

Laminin V Alpha6Beta4-integrin BP antigen

Proliferation markers

Ki67

REFERENCES

Adding the immune component in reconstructed human skin and eye epithelia models.

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Cellular mechanistic investigation on antigen delivery by Viaskin® patchfor epicutaneous immunotherapy with reconstructed human epidermis including Langerhans cells (SkinEthicTM RHE-LC). Dhelf V, Dioszeghy V, Ligouis M, Mondoulet L, Pellevoisin C, Sahuc F. Poster

Reconstructed Human Pigmented Epidermis



 0.5 cm^2



 4 cm^2



HTS 24-well plate 0.33 cm²



HTS 96-well plate 0.11 cm²



APPLICATIONS



Pigmentation



Depigmentation



UV Exposure / Phototoxicity / Photoprotection





Reconstructed Human Pigmented Epidermis



 0.5 cm^2



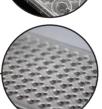
 4 cm^2



HTS 24-well plate 0.33 cm²



HTS 96-well plate 0.11 cm²



APPLICATIONS



Pigmentation



Depigmentation



UV Exposure / Phototoxicity / Photoprotection







Reconstructed Human Pigmented Epidermis



NAME

SkinEthic™ RHPE / Reconstructed Human Pigmented Epidermis

DESCRIPTION

The SkinEthic[™] RHPE model is composed of normal human keratinocytes cultivated in the presence of melanocytes of phototype II, IV or VI, localized in the basal layer.

The different tanning degrees of these constructs correspond macroscopically to 3 different phototypes of human skin.

SPECIFIC MARKERS

Melanocytes localized in the basal cell layer interspersed with basal cell keratinocytes.



Melanin distribution in the basal layer

▶ Ki 67

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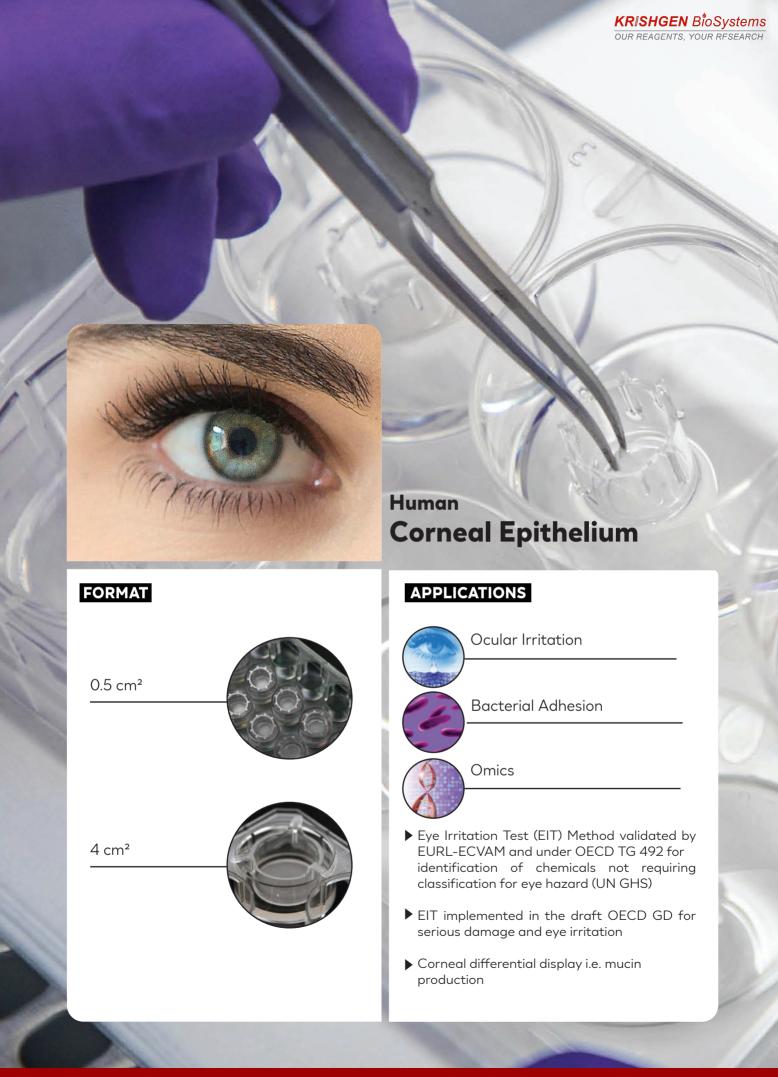
Reconstructed Human Pigmented Epidermis (RHPE): an in vitro model for the evaluation of melanogenesis. Sahuc F. SOFW Magazine.

Sepicalm VG, a new skin lightening enable to modulate melanogenesis-related genes and to prevent UV-induced pigmentation thanks to its soothing properties.

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Human **Corneal Epithelium**



NAME

SkinEthic[™] HCE / Human Corneal Epithelium

DESCRIPTION

The SkinEthic[™] HCE model is composed of transformed human corneal keratinocytes cultivated on an inert polycarbonate filter at the air liquid interface in a chemically defined medium.

The reconstructed tissue forms a stratified and well organized epithelium which is structurally, morphologically and functionnally similar to the human cornea with presence of basal, wing and mucus production cells.

SPECIFIC MARKERS

Differentiation markers:

- Keratin
- **CD44** Hemidesmosomes





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In vitro assessment of eye irritancy using the Reconstructed Human Corneal Epithelial SkinEthic™ HCE model: Application to 435 substances from consumer products industry. Cotovio J et al. Toxicology In Vitro, 24, 523-537.

Are coffee silverskin extracts safe for topical use? An in vitro and in vivo

Alves RC, Beatriz M, Ferreira M, Helena Amaral M, Oliveira PP, Pereira C, Pimentel FB, Rodrigues F, Sarmento B. Elsevier.



are epidermal models on an innovative insert. Epidermal or mucosal reconstructed on this insert are histologically similar to in vivo human epidermis and mucosa.



Human Gingival Epithelium

APPLICATIONS









Human **Oral Epithelium**



NAME

SkinEthic™ HOE / Human Oral Epithelium

DESCRIPTION

The SkinEthic[™] HOE model is composed of TR146 cells (derived from a squamous cell carcinoma of the buccal mucosa) cultivated on an inert polycarbonate filter at the air liquid interface in a chemically defined medium.

This model forms an epithelial tissue devoid of stratum corneum. resembling histologically to the mucosa of the oral cavity.

SPECIFIC MARKERS

Differentiation markers:

Keratin 6 Keratin 16

- **CD44**
- Ki67





REFERENCES

Evaluation of an oral care product safety screening program utilizing the in vitro SkinEthic Human Gingival Epithelium (RHG) and Oral Buccal (RHO) models. Wurzburger L, Kazmi P, Re T, Alonso A, Bertino B, Barnes N, de Brugerolle de Fraissinette A, Hilberer A, Raabe H, Wilt N, Srinivasan V. SOT.

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Oesophageal Epithelium

FORMAT

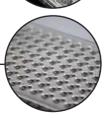
 0.5 cm^2



HTS 24-well plate 0.33 cm²



HTS 96-well plate 0.11 cm²



APPLICATIONS

Oesophageal Irritation



Bacterial Adhesion





Human Oesophageal Epithelium



NAME

SkinEthic™ HO2E / Human Oesophageal Epithelium

DESCRIPTION

The SkinEthic[™] H02E model is a human oesophageal epithelium composed of immortalized cell line Kyse 510, cultivated on an inert polycarbonate filter at the air liquid interface in a chemically defined medium.

This model forms an epithelial tissue devoid of stratum corneum, resembling histologically to the outer cell layers of the human oesophagus.

SPECIFIC MARKERS

Differentiation markers:

- Keratin 6
- Keratin 13





REFERENCES

Influence of voriconazole and fluconazole on reconstituted multilayered oesophageal epithelium infected by Candida albicans. *J.Bernhardt, H.Bernhardt, M.Knoke, K.Ludwig. Mycoses, 47, 7, p.330, October 2004.*

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In Vitro Modelling of Barrier Impairment Associated with Gastro-Oesophageal Reflux Disease (GERD). Marisa Meloni, Paolo Buratti, Francesco Carriero, Laura Ceriotti. Clinical and Experimental Gastroenterology 2021.

Protective Mechanisms of Liquid Formulations for Gastro-Oesophageal Reflux Disease in a Human Reconstructed Oesophageal Epithelium Model. Laura Ceriotti, Paolo Buratti, Enrico Stefano Corazziari, Marisa Meloni. 2022.

Oesophageal Epithelium

FORMAT

 0.5 cm^2



HTS 24-well plate 0.33 cm²



HTS 96-well plate 0.11 cm²



APPLICATIONS

Oesophageal Irritation



Bacterial Adhesion





Human Oesophageal Epithelium



NAME

SkinEthic[™] HO2E / Human Oesophageal Epithelium

DESCRIPTION

The SkinEthic[™] H02E model is a human oesophageal epithelium composed of immortalized cell line Kyse 510, cultivated on an inert polycarbonate filter at the air liquid interface in a chemically defined medium.

This model forms an epithelial tissue devoid of stratum corneum, resembling histologically to the outer cell layers of the human oesophagus.

SPECIFIC MARKERS

Differentiation markers:

- Keratin 6
- Keratin 13





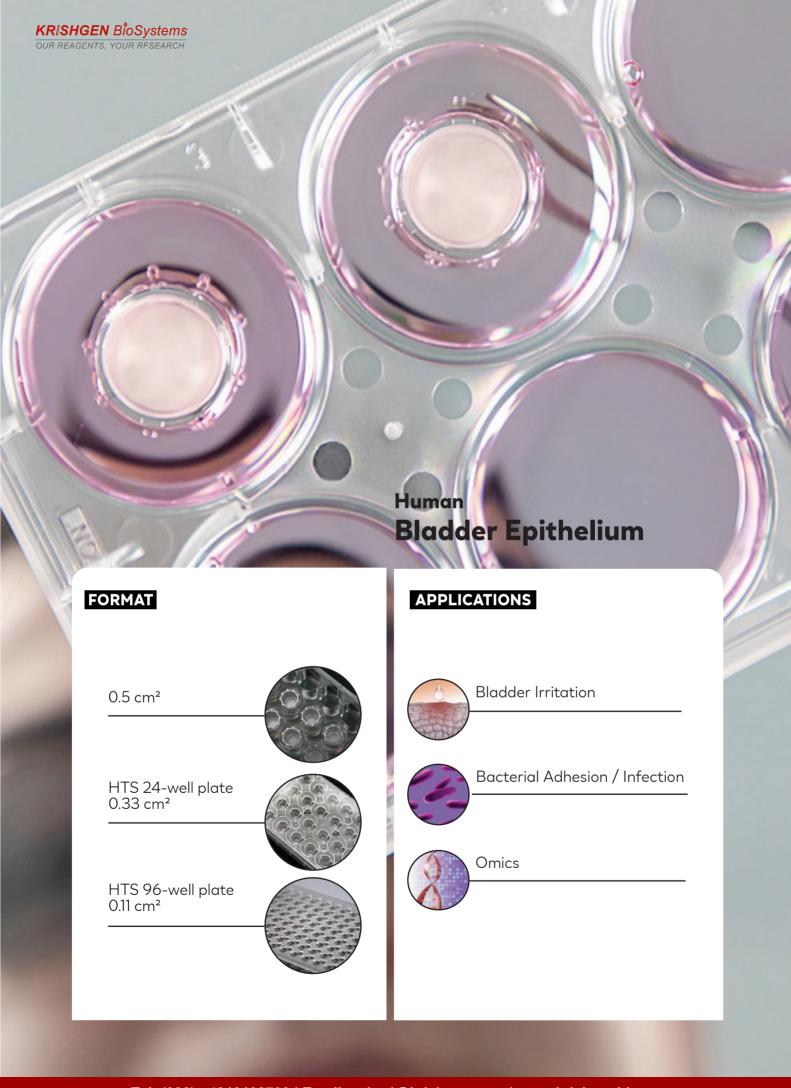
REFERENCES

Influence of voriconazole and fluconazole on reconstituted multilayered oesophageal epithelium infected by Candida albicans. *J.Bernhardt, H.Bernhardt, M.Knoke, K.Ludwig. Mycoses, 47, 7, p.330, October 2004.*

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Protective Mechanisms of Liquid Formulations for Gastro-Oesophageal Reflux Disease in a Human Reconstructed Oesophageal Epithelium Model. Laura Ceriotti, Paolo Buratti, Enrico Stefano Corazziari, Marisa Meloni. 2022.







Human Bladder Epithelium



NAME

SkinEthic™ HBE / Human Bladder Epithelium

DESCRIPTION

The SkinEthic[™] HBE model is a human bladder epithelium composed of immortalized cell line RT-112 (urinary bladder transitional cell carcinoma), cultivated on an inert polycarbonate filter at the air liquid interface in a chemically defined medium.

This model forms an epithelial tissue devoid of stratum corneum, resembling histologically to the outer cell layers of the human bladder.

SPECIFIC MARKERS

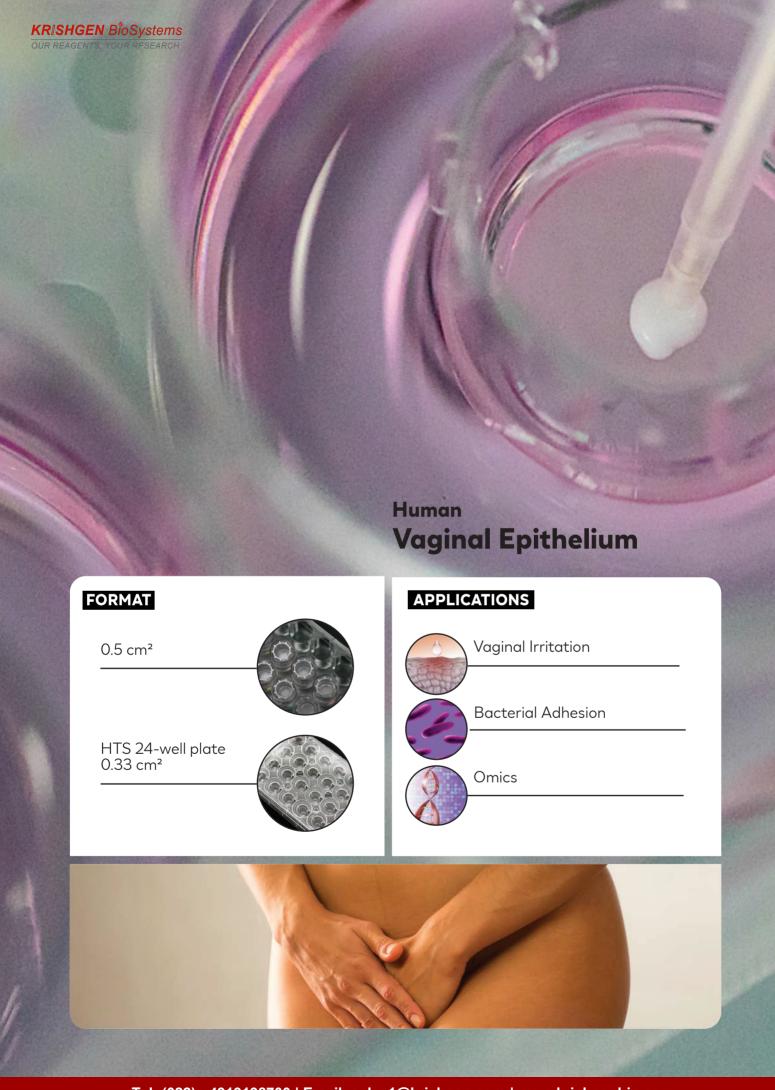
Differentiation markers:

- Keratin 17
- ► Keratin 20
- ▶ CD44











Human **Vaginal Epithelium**



NAME

SkinEthic™ HVE / Human Vaginal Epithelium

DESCRIPTION

The SkinEthic™ HVE model is composed of A431 cells (derived from a vulval epidermoid carcinoma) cultivated on an inert polycarbonate filter at the air liquid interface in a chemically defined medium.

This model is histologically similar to the in vivo vaginal mucosa.

SPECIFIC MARKERS

Differentiation markers:

- Keratin
- Involucrin





REFERENCES

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Quantitative expression of the Candida albicans secreted aspartyl proteinase gene family in human oral and vaginal candidiasis.

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Infect Immun 73 (5): 2758-2765.

The secreted aspartyl proteinases sap1 and sap2 cause tissue damage in an in vitro model using vaginal candidiasis using reconstituted human vaginal epithelium. Schaller M, Bein M, Korting HC, Baur S, Hamm G, Monod M, Beinhauer S and Hube

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